
Complete nucleotide sequence of the nonstructural protein genes of Semliki Forest virus

Kristiina Takkinen

Recombinant DNA Laboratory, University of Helsinki, Valimotie 7, SF-00380 Helsinki, Finland

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ABSTRACT

The nucleotide sequence coding for the nonstructural proteins of Semliki Forest virus has been determined from cDNA clones. The total length of this region is 7381 nucleotides, it contains an open reading frame starting at position 86 and ending at an UAA stop codon at position 7379-7381. This open reading frame codes for a 2431 amino acids long polyprotein, from which the individual nonstructural proteins are formed by proteolytic processing steps, so that nsP1 is 537, nsP2 798, nsP3 482 and nsP4 614 amino acids. In the closely related Sindbis and Middelburg viruses there is an opal stop codon (UGA) between the genes for nsP3 and nsP4 (1). Interestingly, no stop codon is found in frame in this region of the Semliki Forest virus 42S RNA. In other aspects the amino acid sequence homology between Sindbis, Middelburg and Semliki Forest virus nonstructural proteins is highly significant.

INTRODUCTION

Semliki Forest virus (SFV) belongs to the alphaviruses and is, together with Sindbis virus, the best characterized member of this virus group. The viral nucleocapsid, composed of a capsid protein and a RNA genome, is surrounded by a lipid membrane with viral glycoproteins E1, E2 and E3 (for reviews see 2, 3). The genome of SFV, the 42S RNA, is a single stranded RNA of positive polarity with a 5' terminal cap structure and poly(A) region at the 3' end (4, 5, 6). In the early phase of infection the 5' two thirds of the genome are translated as a polyprotein to the nonstructural (ns) proteins, required for the replication of viral RNA. In the first step of viral replication, a full length minus strand is synthesized, which then serves as a template for new plus 42S RNA synthesis (7, 8). The mRNA for the four viral structural proteins, a subgenomic 26S RNA, is transcribed starting at an internal initiation site on the minus strand 42S RNA template (4, 9). The 26S RNA is also translated as a polyprotein which is

proteolytically processed to the four structural proteins. Temperature-sensitive mutants of Sindbis virus, which have defects in the RNA synthesis fall into four complementation groups, suggesting that four different proteins are involved in alphavirus RNA replication (10, 11, 12).

The translation order of the ns proteins of SFV, nsP1 (ns70)-nsP2(ns86)-nsP3(ns60)-nsP4(ns72), has been determined by using salt synchronized protein synthesis and specific labeling conditions (14, 15). The present nomenclature of these proteins, nsP1, nsP2, nsP3 and nsP4 reflects to the order of translation (13), the previous ns70, ns86, ns60 and ns72 to their apparent molecular weights. The ns proteins of SFV were first identified in cells infected with a temperature-sensitive mutant, ts-1, which turned out to be an over producer of the ns proteins (16, 17). Various short lived intermediate cleavage products are found in cells infected with ts-mutants. Two intermediate cleavage products of the 250 kd ns polyprotein, ns155 and ns135, are found in cells infected with ts-1 containing the sequences of nsP1 plus nsP2 and nsP3 plus nsP4, respectively (18). Another precursor ns220, which is synthesized together with the entire polyprotein ns250 in cells infected with ts-4 or ts-6, is processed to nsP1, nsP2 and nsP3 (19, 20). The cleavage intermediates accumulate in mutant infected cells due to impaired proteolytic processing of the primary translation product (21). All of them can be identified in cells infected with wild type SFV, but in much smaller quantities. The synthesis and processing of the ns proteins of Sindbis virus is different, because of the stop codon between nsP3 and nsP4 (1). The major precursor polyprotein (p230) of Sindbis is processed to nsP1, nsP2 and nsP3. Only minor amounts of the full length ns polyprotein, from which the nsP4 is generated, is synthesized by read through of the stop codon (1, 13).

The proteolytic processing sites of the ns polyprotein of SFV have been determined by direct NH₂-terminal amino acid sequence analysis of radiolabeled ns proteins (22, Kalkkinen et al., to be published). This allows to localize the genes for the individual ns proteins in the nucleotide sequence presented in this paper. The 26S RNA region of SFV has been cloned and sequenced earlier

(23, 24), so now the total structure of the genome of SFV is known. Nucleotide sequence data from other alphaviruses is also wide; at present the complete nucleotide sequence of the genome of Sindbis virus has been determined (13, 25) and parts of the ns genomic region of Middelburg virus has been sequenced (1). This sequence information of the ns protein genes provides possibilities to search for conserved domains of the ns proteins with important functions in the RNA replication of alphaviruses.

MATERIALS AND METHODS

Enzymes and reagents

Enzymes were obtained from commercial sources, isotopes $[\gamma\text{-}^{32}\text{P}]$ ATP (3000 Ci/mmol), $[\alpha\text{-}^{32}\text{P}]$ dNTP (3000 Ci/mmol), $[\alpha\text{-}^{32}\text{P}]$ ddATP (3000 Ci/mmol) and $[\alpha\text{-}^{35}\text{S}]$ dATP (~ 600 Ci/mmol) from Amersham. The oligonucleotide primer (15-mer) used for the direct RNA sequencing was synthesized in the Beckman Application Laboratory.

cDNA synthesis and cloning

Viral 42S RNA was isolated from purified virions disrupted with 2 % SDS and was purified on 15-30 % sucrose gradient as described (26). Restriction enzyme fragments of about 100-200 bp in length were isolated as primers for the cDNA synthesis by linear 2.5-10 % polyacrylamide gradient gels (27) from the cDNA clones in pBR322. The desired fragments were eluted by diffusion (28) or electrophoresed onto DEAE-cellulose membranes (NA-45 Schleicher&Schüll) and recovered with 50 mM TRIS-HCl pH 7.5, 1 M NaCl and precipitated with ethanol. 5 μ g of viral RNA was ethanol precipitated with 5-10 fold molar excess of primer, washed with 70 % ethanol, dried and dissolved in 5 μ l of water. The template-primer mixture was denatured at 90°C for 3 min and annealed at 60°C for 20 min in 50 mM TRIS-HCl pH 8.3, 0.6 M KCl. The reaction mixture (100 μ l) for the first strand synthesis contained: 50 mM TRIS-HCl pH 8.3, 10 mM MgCl_2 , 140 mM KCl, 5 mM DTT, 500 μ M dATP, dGTP, TTP, 250 μ M dCTP, 30 μ Ci of $[\alpha\text{-}^{32}\text{P}]$ dCTP, 150 U RNase inhibitor and 70-100 U reverse transcriptase (Life Science or Promega Biotec). The synthesis was at 42°C for 60 min, and was stopped by adding EDTA to 20 mM. The cDNA was extracted with phenol and precipitated with ethanol from 2 M ammonium acetate. The second strand synthesis was done either according to Gubler and Hoffman (29) or by the following procedure: The RNA

was hydrolyzed at 50°C for 30 min in 65 mM NaOH and the nucleotides were removed by gel filtration on Biogel P-30 (Biorad) and ss cDNA was recovered with ethanol precipitation. The second strand was synthesized with reverse transcriptase as above, omitting RNase inhibitor and $[\alpha\text{-}^{32}\text{P}]$ dCTP and the concentration of dCTP was 500 μM . The reaction was stopped and cDNA recovered as before. The ds cDNA was then treated with 1 U of S1-nuclease in 50 μl of 50 mM NaCl, 30 mM Na-acetate pH 4.6, 1 mM ZnCl_2 . The ds cDNA was size selected on a 2 ml column of Sephacryl S-1000 (Pharmacia). The 3' termini of the cDNA were elongated with dCTP and terminal transferase to give homopolymeric tails (10-30 residues) and vector pBR322, cleaved with PstI, was similarly tailed with dGTP (30), the cDNA was annealed to a molar amount of vector. Some clones were prepared by ligating SalI cleaved cDNA into SalI site of pBR322. Transformation into *E. coli* HB101 was according to Mandel and Higa (31). Small scale plasmid DNA was prepared (32) from colonies which had the correct antibiotic resistance pattern and were then assayed for colinearity with viral 42S RNA using S1-nuclease test (33) and large scale plasmid DNA was prepared as previously described (33).

DNA sequencing

The nucleotide sequences of the cDNA clones were determined by Maxam and Gilbert (28) or Sanger dideoxy (34) methods. The restriction enzyme maps of the inserts were constructed by partial digestions of end labeled insert fragments (35) or double digestions. For chemical sequencing the 5' ends of the fragments were first dephosphorylated with calf intestinal alkaline phosphatase and then labeled with T4-polynucleotide kinase and $[\gamma\text{-}^{32}\text{P}]$ ATP (27). The 3' ends were labeled either with Klenow fragment and appropriate $[\alpha\text{-}^{32}\text{P}]$ dNTP (36) or by terminal transferase and $[\alpha\text{-}^{32}\text{P}]$ ddATP (30). Labeled DNA fragments were digested with secondary restriction enzyme and isolated from polyacrylamide gradient gels or strand separated (28) and purified from contaminating polyacrylamide by mini DE 52 cellulose columns (37). DNA sequences were analyzed on 20 %, 8 % and 6 % polyacrylamide gels (38).

For the dideoxy sequencing, insert specific restriction fragments were isolated by polyacrylamide gradient gels, ligated to

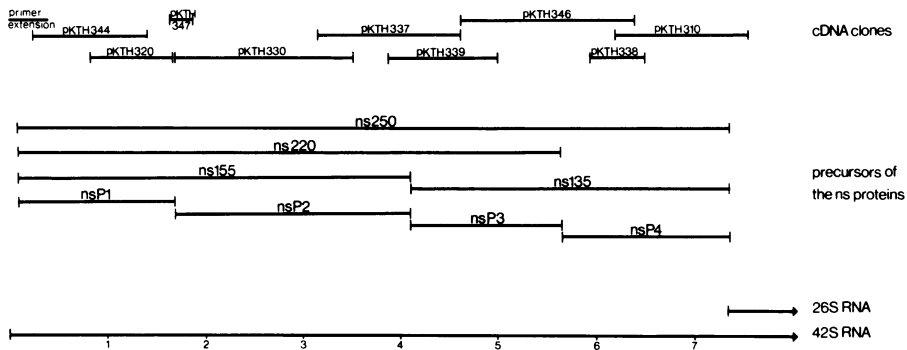


Figure 1 Schematic representation of the nine cDNA clones in the ns protein coding region of SFV used in the nucleotide sequence determination. The nucleotide sequence of the 5' end of the genome has been determined earlier by primer extension (37). The ns polyprotein, the intermediate and the final proteolytic cleavage products are in the middle. The bottom line indicates the scale in kilobases and shows the location of the 5' end of 26S RNA (45).

dephosphorylated M13 vectors, and transfected to JM103 (39). The sequences from recombinants with complementary strands were determined (39) using $[\alpha\text{-}^{35}\text{S}]$ dATP as the label (40).

For the direct RNA sequencing about 20 ng of the primer was annealed with 1-2 μg of viral RNA and sequenced with reverse transcriptase (41) using $[\alpha\text{-}^{35}\text{S}]$ dATP as the label.

RESULTS AND DISCUSSION

Cloning and sequencing the ns protein genes of SFV

The 3' third of the 42S RNA genome of SFV, the 26S RNA region, coding for the four viral structural proteins has been cloned and sequenced earlier (23, 24). A restriction fragment from the 5' end of this cDNA clone (kindly provided by dr. H. Garoff) was used as a primer to synthesize cDNA for the ns protein coding region. The cDNA was cloned into pBR322 by dC:dG tailing. Restriction fragments from two clones obtained, pKTH310 and pKTH320 (Fig. 1), were used as primers for further cDNA synthesis. After two cloning steps about 85 % of the ns protein coding region was covered in three blocks. The restriction enzyme map data of these cDNA clones was the basis for the third cloning step. The two gaps were flanked by SalI sites, thus to obtain clones for these

regions the ds cDNA was cleaved with Sall and cloned into the Sall site of pBR322. The ns protein coding region was covered in nine overlapping clones (Fig. 1) except the 250 nucleotides from the 5' end, which has been determined earlier by primer extension of viral RNA (37) and the 22 5' terminal nucleotides by direct RNA sequencing of the antigenome (42). To avoid cloning artefacts the colinearity of the cDNA clones with 42S RNA was determined with S1-nuclease (33). The nucleotide sequence of the cDNA clones was determined by the Maxam and Gilbert (28) and Sanger dideoxy (34) methods. The nucleotide sequence data was analyzed by the computer programmes of Staden (43) and Peltola (44).

Complete nucleotide sequence of the nonstructural region

The nucleotide sequence of the ns protein genes and the deduced amino acid sequences are shown in Figure 2. The total length of the 42S RNA is 11442 nucleotides plus the poly(A) tail, which is approximately 80-90 nucleotides in length (5, 6). The base composition of the 42S RNA is 26.7 %A, 20.1 % U, 27.0 % G and 26.2 % C. The molecular weight of the 42S RNA is 3.95×10^6 daltons in the sodium form, without the poly(A) tail. There is an open reading frame in the ns region starting with AUG at nucleotide 86 and ending at a termination codon UAA at nucleotides 7379-7381, coding for a 2431 amino acids long polyprotein. The two other reading frames are blocked with multiple stop codons, 118 and 132, respectively. The total length of the 26S RNA, mRNA for the structural proteins, is 4074 nucleotides, from which the 5' noncoding region is 51 nucleotides, giving an overlap of 13 nucleotides with the ns coding region (45). Translation of the structural polyprotein starts 38 nucleotides from the stop codon of the ns region in a different reading frame from that used for the ns proteins (45).

Nucleotide sequence between the genes of nsP3 and nsP4

The nucleotide sequence of the ns protein genes of Sindbis and Middelburg viruses revealed an opal stop codon UGA six amino acids upstream from the NH₂-terminal amino acid of nsP4 (1). In the sequence of SFV at the same position there is an arginine codon CGA, showing a change from U to C when compared to Sindbis and Middelburg. The sequence downstream from this codon is highly conserved especially between SFV and Middelburg, but shows little

no: P2

V L T C F R G W A K Q L Q L D T R B H E V N T A A A S Q G L T R E O V T A V R Q
C T G T T A A C A T G T C T T G C G A G G T G G G C A A G C A G T C C A G T T G G A C T A C C T G G A C A C G A A G T C A T G C A G C A G C A G C A T C T C A G G G C T C A C C C G C A A A G G G T A T A C G C G T A A G G C A
2770 2780 2790 2800 2810 2820 2830 2840 2850 2860 2870 2880

K V H E N F L Y A P A S E R V H V L L T R T E D R L V W K T L A G D P W I E V L
G A A G G T G A A T G A A A T C C T T G T A T G C C C T G C T G C G A G C A C G T G A A T G T A C T G C T A G C G C A C T G A G G T A G G C T G G T G T G G A A G C C G G C C G G A A C C T G G A T T A A G G C C T
2890 2900 2910 2920 2930 2940 2950 2960 2970 2980 2990 3000

S N I F Q G H F T A T L E W Q S E H D K I N K V I E G P A A P V D A F Q N K A
A T C A A A C A T T C C C A G G G T A A C T T A C T G G C C A C A T T G G A A G A A T G C C A A G A A C A C G A C A A A A T A A T G A A G G T A A A G A A G C C G C T G C C A G A A C A A A G C
3010 3020 3030 3040 3050 3060 3070 3080 3090 3100 3110 3120

H V C W A K S L V F V L D T A G I R L T A E W S T I I T A F E D R A T S P V
G A A C T G T G T G T G G G C A A A A C C G T G C T T C T G G A C A C T G C C G A A T C A S A T T G A C A G C A G A G G A T G G A G C A C C A T A A T T A G A G A G A C A G A G C T T A C T C C C A G T
3130 3140 3150 3160 3170 3180 3190 3200 3210 3220 3230 3240

V A L H E I C T E Y Y Q V D L D S G L F S A P E V S L Y E N H N W D H R P Q G
G G T G G C T T G A A T G A A T T T G C A C C A A G T A C T A T G A A T T G A C T G G A C A G T G C C T G T T T C T G C C C C A A G G T G C C C T G T A T T A C G A A A C A A C A C T G G G A T A A C A G A C T G G T G
3250 3260 3270 3280 3290 3300 3310 3320 3330 3340 3350 3360

R H Y G F N A A T A A R L E A R N T F L K G Q W E T G K Q A V I A E R R E I Q P L
A A G A G T G A T G A T G A A T G C C C A G C A G C T G C C A G C T G G A A G T A G A C A T A C T T C T G A A G G G C A G T G G C A T A C C G G C A A G C A G A G T A T T C G C A G A A A A G T A A G A G G T C C A C C C T
3370 3380 3390 3400 3410 3420 3430 3440 3450 3460 3470 3480

S V L D H V I P I N R R L P H A L V A E Y E T V E G S E R V E W L V H K V R G Y H
T T C T G T G C T G A C A A T G T A A T T C T A T C A A C C G A C G T G C C G C A C G C C T G T G T G C T G A G T A C A A G A C G G T A A A G C A G T A G G T T A A G T G G C T G G T C A A T A A A G T A A G A G G T A C C A
3490 3500 3510 3520 3530 3540 3550 3560 3570 3580 3590 3600

V L L V S E Y H L A L P R R R V T W L S P L E V T G A D R C Y T D L S L G L P A D
C T C C T G C T G T G A G T A C A A C T G C T T T G C C G A C G C A G G C A C T T G G T T C A C C G C T A A T G T C A G A G C C C G A T A G G T G C T A C G A C T A G T T T A G A G A T C C C G C T G A
3610 3620 3630 3640 3650 3660 3670 3680 3690 3700 3710 3720

A G R F D L F V F W I N T E F R I E N Y Q C V D H A N K L Q H L G G D A L R L
C C C G G C A G T T G C A T T G G T T T G T G A A C A T T C A C A G G A A T C C A C A C T A C C A G C A G T G T G T C A C C A C C C A A G G T G C C A G A T G C T T G G G G A G A T G C G C T A C G A C T
3730 3740 3750 3760 3770 3780 3790 3800 3810 3820 3830 3840

L K P G G I L N R A Y O G Y A D E I S E A V V S S L S R E F S S A R V L R P D C V
G T A A A A C C C C G G C G A C T C G T A T G A G A G C T T A C G G A T A C C C G A T A A A T C A G C G A A G C G T T G T T C T C T C T A A G C A G A A G T T C T C G T C G C A A G A T G T T G C G C C G A T T G T G T
3850 3860 3870 3880 3890 3900 3910 3920 3930 3940 3950 3960

T S H T E V F L L F S F D D N G K R P S T L H Q N H T K L S A V Y A G E A N H T
C A C C A G C A T C A G A A T G T T C T G T G T T C C A A C T T G A C A C G A A A G A G A C C C T A C G C T A C A C C A G A T A A T A A G C A A G C T G C T G T A T G C C G A A A G C C A T G C A C A C
3970 3980 3990 4000 4010 4020 4030 4040 4050 4060 4070 4080

A G C A P S Y E V E R A D I A T C T E A A V V H A A H A R G T V D G V C R A V
G C C G G G T G C C A C C A T C C T A C A G A G C A G A C A C A T G C C A G A A G G G C T G G T G A T C C A G C C C G T G G A A C T G T A G G G G A T G C C T A T G C A G A G G C C T
4090 4100 4110 4120 4130 4140 4150 4160 4170 4180 4190 4200

A K E W P S A F E G A A T P V G T I E T V H C G S T P V I H A V A P H F S A T T
G C G A A G A A A T G C C G T C A G C T T T A A G G A G C A A C A C C A G T G G G C A C A A T A A A C A G T C A T G T G C G G C T G T A C C C C G T C A T C C A C G C T G A G C C G A T T C T C T G C C A C A C
4210 4220 4230 4240 4250 4260 4270 4280 4290 4300 4310 4320

E A E G D R E L A A V T R A V A A E V H R L S L S S V A I P L L S T G V F S G G
T G A A G C G A A G G G G C C G A A T T G G C G C T G T C A C C G G G C A G T G G C C G G A A T A A C A G A C T G C T A C T G A G C A G T A G C A T T C C C G C T G T G C C A G A G A G T T C A G C G G G C A
4330 4340 4350 4360 4370 4380 4390 4400 4410 4420 4430 4440

R D R L Q Q S L N H L F T A R D A T D A D V T I T C R D K S W E E K I Q E A I D
A A G A T A A G C T G C A G A C A T C C C T C A C C A T C T A T T C A C A G C A A T G A G C C A G C A G A G C T G A C T G A C A G A C A A A A A T C C A G A A G C A A T C A T T G A
4450 4460 4470 4480 4490 4500 4510 4520 4530 4540 4550 4560

H R T A V E L L H D D V E L T T D L V R V H P D S S L V G R E K G Y S T T D G S L
C A T G A G G A C G G C T G G A G T T C T A A T G A T C A T G G A G C T G A C C A C A G A C T T G T G A G A G T G C A C C C G A C A G C A G C C T G T G G G T G R E A A G G T A C A G T A C C A G A C G G T C C T
4570 4580 4590 4600 4610 4620 4630 4640 4650 4660 4670 4680

T S Y F E G T K F H Q A A I D H A R I L T L W P R L Q E A H E R I C L T A L G E
G T A C T C T A T T T G A A G G T A C A A A T T C A A C A G C G T G C T A T T A T G C A G A T A C T G A C O T T G G G C C A G A C T G C A G A G G C A A C G A A G A T G C T A T A C C A C G C T G G C G A
4690 4700 4710 4720 4730 4740 4750 4760 4770 4780 4790 4800

T N D H I G S K C P V H D S D S T S T P P R T V P C L R T A N T A S E R I A R L R
A C A A T G A C A A C A T C C G A T C C A A A T G C C G G T G A A G A T T C C G A T T C A T C A A C A C T C C C A G A G A C A G C C T G C T G T G C C T A C C A A T G A C A G A A A C G A A T C C C C C T T A G
4810 4820 4830 4840 4850 4860 4870 4880 4890 4900 4910 4920

S H O V K S H V C S S P F L P E Y H V D G V Q E V E C E R V L L F D P T V P S
G T C A C A A G A T A A A G A T G G T G G T G C T A C T T T C C C T C C G A A A T A C C A T G A T G A G T G G G T G C A G A A G T A A A G T G C G A A G A G T T C T C T T G C A C C A G C A G T A C C T
4930 4940 4950 4960 4970 4980 4990 5000 5010 5020 5030 5040

V V S P R K Y A A S T T D H S D R S L R G F D L D W T T D S S S T A S D T H S L
A G T G G T A T G C C G G A A G T A T C C G C A T C A C A G C G G A C A C T C A G A T C G G T C G T A C G A G G G T T G A C T T G G A C T G G A C C A C C A G C T G C T T C C A C T G C A G C A G A T A C C A T G C T C
5050 5060 5070 5080 5090 5100 5110 5120 5130 5140 5150 5160

P S L Q S C D I D S I Y E P N A P I V V T A D V H P E P A G I A D L A A D V H P
A C C A G T T G C A G T C G T G A C A T C G A C T G A T C A C G A G C A A T G G C T C C C A T A G T A G T G A C G C T G A C G T A C C C T G A A C C C G A G C A G C C G G A C T G G C G G A G A T G T G C A C C
5170 5180 5190 5200 5210 5220 5230 5240 5250 5260 5270 5280

E P A D H V D L E H P I P P F R P K R A A Y L A S B A A E R P V P A P H K P T P
T G A A C C C C G A C C A T C C C T G A G A C C G A T T C T C C A C C G C G C C G A A G A G C G C A T A C T G C C T C C C G C C G G C G G A C C G G T G C C G C C G G A A A G C C A G C A C
5290 5300 5310 5320 5330 5340 5350 5360 5370 5380 5390 5400

A P R T A F R N K L P L T F G D F D E H E V D A L A S G I T F G D F D D V L R L
T G C C C A A G A C T G C G T T T A G G A A C A G C T G C T T T G A C G T T G C G G C A C T T T G A C G A C A G C A G G T C A G T C G T G G C C T C C G G A T T A C T T T C G G A G A C T T C G A C A G C C T G C G A C T
5410 5420 5430 5440 5450 5460 5470 5480 5490 5500 5510 5520

homology upstream (Fig. 3a). To rule out the possibility of a cloning artefact at this region, the DNA sequence of the cDNA clone was confirmed by direct RNA sequencing from a specific

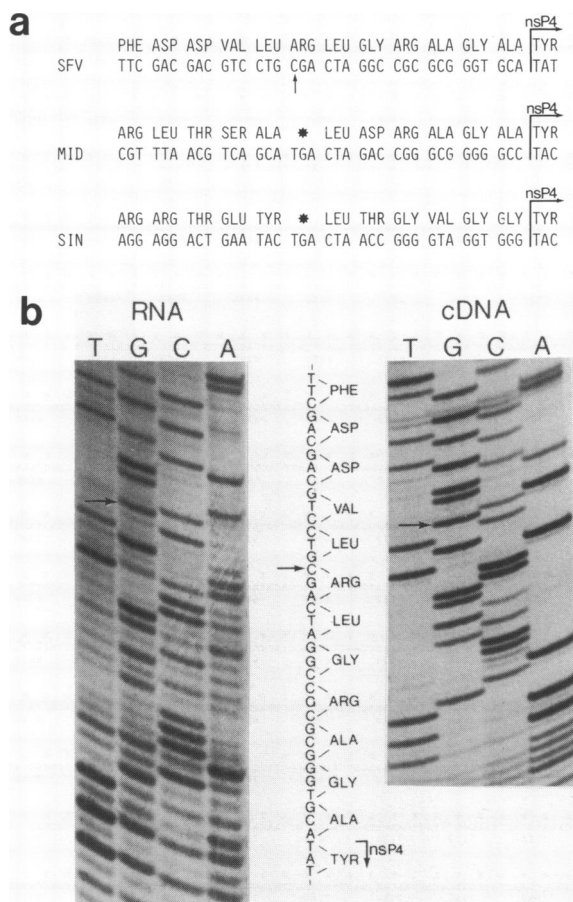


Figure 3. a. Comparison of the nucleotide sequences (written in DNA form) of SFV, Middelburg (MID) and Sindbis (SIN) viruses between the genes of nsP3 and nsP4. The opal stop codon (TGA) of Middelburg and Sindbis viruses is marked with a star and the arginine codon (CGA) at the same position in the genome of SFV with an arrow.

b. Sequence analysis of the SFV RNA and cDNA clone from the same region. Both gels show the complementary sequence of the coding strand written in the middle. The C residue in the arginine codon is marked with an arrow.

oligonucleotide primer, which located about 40 nucleotides downstream from the starting point of nsP4. The result of the direct RNA sequencing was identical with the sequence obtained from the cDNA clone, although there was background at some positions, showing that the nucleotide at this position in the

genome of SFV is C (Fig. 3b). This finding is compatible with earlier results obtained from the translation studies of the ns proteins of SFV (14, 15, 21, 46), which have shown that nsP4 (ns72) can clearly be detected in cells infected with ts-1 mutant, an overproducer of the ns proteins, and also with the wild type SFV. Furthermore, Keränen and Ruohonen (15) identified the carboxyterminal translation product of SFV *in vivo* using 30 sec pulses of ^{35}S -methionine. The only carboxyterminal ns protein detected was nsP4 together with its immediate precursor nsl35. The existence of nsP4 in Sindbis virus infected cells has been shown only by immunoprecipitation with an antibody prepared against a synthetic COOH-terminal peptide of nsP4 (1). The different expression level of the nsP4 between these virus is, however, surprising, because it is the most conserved ns protein between SFV and Sindbis, indicating an important role for it in the RNA replication.

Deduced amino acid sequences

For the localization of the individual ns protein genes in this nucleotide sequence the partial aminoterminal amino acid sequences of radiolabeled nsP2, nsP3, nsP4 and nsl35 were determined (22, Kalkkinen et al. to be published). The NH_2 -terminus of nsP1 and its precursor nsl55 is blocked, but the initiation dipeptide for the ns polyprotein has been shown to be met-ala (47). The AUG codon at position 86-88 is followed by an alanine codon and the only open reading frame in the ns region starts at this point, thus it is quite obvious that nsP1 starts at this position. It should, however, be noted that the initiation codon is preceded by two other AUG:s at positions 1 and 8. The proteolytic processing sites yielding the obtained NH_2 -termini are marked with arrows in Figure 2. The proteolytic cleavages in the ns polyprotein of SFV occur between alanine-glycine (AG), cysteine-alanine (CA) and alanine-tyrosine (AY), alanine and glycine are preceding all the cleavage sites. The details of the processing steps are unknown, but based on the accumulation of cleavage intermediate (ns220) and the unprocessed polyprotein by certain ts-mutants it has been suggested that one or more of the ns proteins could be involved in this processing (21).

The molecular weights of the ns proteins calculated from the amino

Table 1. Molecular weights of the nonstructural proteins of SFV

	nsP1	nsP2	nsP3	nsP4
Determined by polyacrylamide gel electrophoresis (Keränen & Ruohonen 1983)	64000	86000	61000	68000
Determined from the nucleotide sequence	59600	88500	52100	68900

acid compositions agree quite well with those determined by gel electrophoresis (Table 1), except in the case of nsP3, which has almost 10000 daltons lower molecular weight than estimated earlier. The carboxyterminus of nsP3 has some interesting features. It has a high proline content and it contains amino acid repeats; two identical octapeptides (ADVHPEPA), tetrapeptides (PAPR) and hexapeptides (TFGD²FD), these are underlined in Figure 2.

Codon usage

The codon usage of the ns region is shown in Table 2 and compared to that used in Sindbis virus (13). In eukaryotic mRNAs and in some eukaryotic viruses, e.g. poliovirus and VSV, there is a low incidence for CpG dinucleotide codons for serine, proline, threonine and alanine (48, 49, 50). The codon usage in the

Table 2. Codon usage in the nonstructural region of SFV and Sindbis virus

	SFV		SIN			SFV		SIN			SFV		SIN			SFV		SIN	
PHE	UUU	32	42	SER	UCU	20	20	TYR	UAU	22	32	CYS	UGU	17	10				
	UUC	47	46		UCC	32	24		UAC	60	47		UGC	44	48				
LEU	UUA	14	13		UCA	23	28	TERM	UAA	1	0	TERM	UGA	0	1				
	UUG	56	39		UCG	29	36		UAG	0	1		UGG	23	22				
LEU	CUU	12	35	PRO	CCU	31	28	HIS	CAU	18	31	ARG	CGU	10	20				
	CUC	22	24		CCC	32	32		CAC	45	31		CGC	25	27				
	CUA	35	33		CCA	29	50	GLN	CAA	16	37		CGA	14	10				
	CUG	76	62		CCG	44	45		CAG	53	53		CGG	10	13				
ILE	AUU	28	42	THR	ACU	33	32	ASN	AAU	21	35	SER	AGU	18	21				
	AUC	50	51		ACC	49	55		AAC	59	62		AGC	33	31				
	AUA	25	31		ACA	49	55	LYS	AAA	66	78	ARG	AGA	45	45				
MET	AUG	53	57		ACG	41	33		AAG	80	88		AGG	43	25				
VAL	GUU	34	34	ALA	GCU	42	38	ASP	GAU	38	50	GLY	GGU	16	25				
	GUC	53	50		GCC	76	76		GAC	105	78		GGC	41	32				
	GUA	37	57		GCA	60	63	GLU	GAA	63	88		GGA	46	51				
	GUG	84	51		GCG	50	43		GAG	73	75		GGG	29	22				

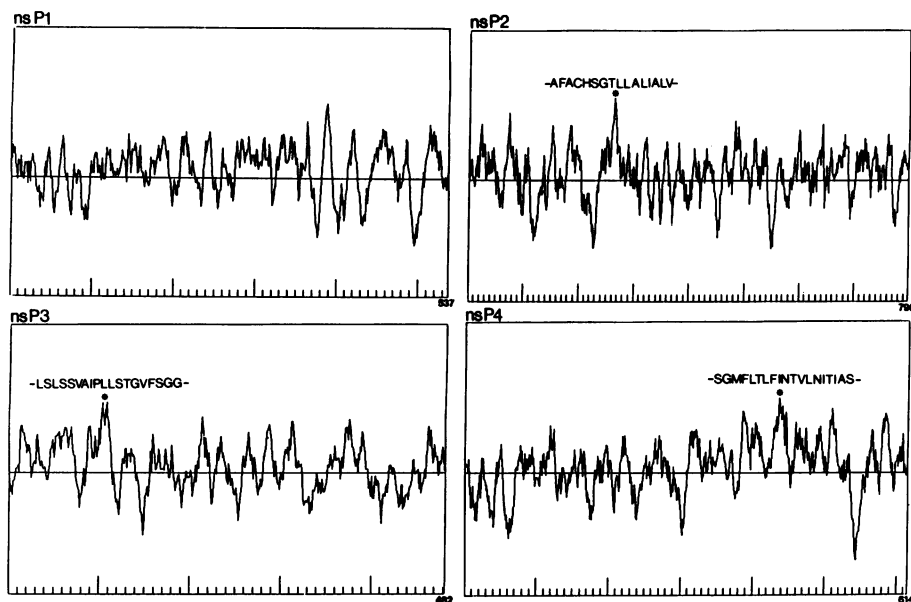


Figure 4. Hydrophobicity plots of the deduced ns proteins of SFV according to Kyte and Doolittle (55) with a search length of seven amino acids. The longest hydrophobic sequences are indicated in one letter code, the scale of each plot shows the length of the protein in amino acids.

structural regions of SFV, Sindbis and Ross River viruses (23, 24, 35, 51) and the ns regions of SFV and Sindbis virus (Table 2) does not show a low CpG incidence, which could reflect the ability of alphaviruses to replicate also in invertebrate hosts. Between SFV and Sindbis virus there is a difference in the codon usage profiles of some amino acids in the ns regions, e.g. in the codons for leucine, valine, tyrosine, histidine and asparagine SFV prefers G or C at the third position in the codon. It is interesting that although the amino acid composition of the ns proteins is extensively conserved, SFV has adapted to a codon usage, in which the codon-anticodon interaction for some amino acids is stronger. Codon usage between different ns proteins and between the structural and the ns regions of SFV are similar.

Hydrophobic features of the ns proteins

The alphavirus replication complex is known to be associated with intracellular membranes (52, 53). NsP1 (ns70), nsP2(ns86) and

nsP4 (ns72) are the proteins that are found from the membrane solubilized replication complex (54). The hydrophobicity plots of the ns proteins according to Kyte and Doolittle (55) are shown in Figure 4.

The carboxyterminus of nsP1 has hydrophobic regions separated by hydrophilic residues, nsP2 has hydrophobic peak around residue 260 and nsP3 contains a stretch of 19 uncharged amino acids starting at residue 99. The carboxyterminus of nsP4 contains a gly-asp-asp (GDD) triplet (nt 6929-6937) surrounded by hydrophobic sequences (underlined with a broken line in Fig. 2). These sequences have been shown to be conserved extensively when poliovirus RNA polymerase sequence was compared with many other putative RNA polymerase coding regions of positive strand RNA viruses of eukaryotes and plants and bacterial viruses (56), representing perhaps a functional domain in the polymerase complex. Similar hydrophobic features are also found in the ns proteins of Sindbis virus (13).

A detailed analysis of the conserved sequences of the ns proteins of SFV and Sindbis virus should lead to some predictions about the functions of these proteins; possible protease function, polymerase function and the regulatory factors involved in the RNA replication event.

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